

Suggested Readings for Project 1

- Skoog D.A.; Holler F.J.; Nieman T.A. *Principles of Instrumental Analysis - 5th Edn.*; Thomson Brooks/Cole: Belmont, CA, USA, 1998.

Spectrophotometric Determination of MnO_4^- and $\text{Cr}_2\text{O}_7^{2-}$ in Their Mixture

Topics to be covered (page: 300-354, ch. 13,14)

- Electromagnetic spectrum
- Beer's Law
- Limitations of Beer's Law
- Absorbing species and electronic transitions
- Instrumentation (components and types of instruments)
- Determination of multi-component mixtures
- Applications of UV-Vis spectrophotometer

Infrared Determination of Benzoic Acid and Salicylic Acid

Topics to be covered (page: 380-428, ch. 16, 17)

- Molecular vibrations
- Vibrational modes and coupling
- Instrumentation
- Sample handling for solids, liquids, and gases
- Qualitative and quantitative analysis
- Reflection spectrometry

If you are using an earlier or newer edition of the book, you must find the related subjects in your textbook.

For additional resources, please check this website.

<http://teaching.shu.ac.uk/hwb/chemistry/tutorials/>

The sub-header you will be concerned is “*Resources for Analytical Science / Molecular Spectroscopy*”. The topics are “Beer's Law”, “UV-Vis. Absorption Spectroscopy”, “Infra-red Absorption Spectroscopy”.

Spectrophotometric Determination of MnO_4^- and $\text{Cr}_2\text{O}_7^{2-}$ in Their Mixture

Purpose: In this experiment, the mixture of MnO_4^- and $\text{Cr}_2\text{O}_7^{2-}$ solutions will be analyzed and determined by UV-Vis spectrophotometer.

Introduction:

Absorption of visible and ultraviolet (UV) radiation is associated with excitation of electrons in an atom or molecule to higher energy states. All molecules can undergo electronic excitation following absorption of light. Because absorption spectra are characteristic of molecular structure, they can be used to qualitatively identify atomic and molecular species. The amount of light, I , transmitted through a solution of an absorbing chemical in a transparent solvent can be related to its concentration by Beers Law:

$$-\log \frac{I}{I_0} = A = \epsilon_{\lambda} bc$$

where I_0 is the incident light intensity, A is the absorbance (a defined quantity, also referred to as the optical density, or OD), b is the cell path length in cm, c is the solution concentration in moles/liter, and ϵ_{λ} is the molar absorptivity, (also referred to as the molar extinction coefficient) which has units of liter/mole/cm (i.e., A is a unitless quantity). Notice that ϵ_{λ} is a function of wavelength, and it is the quantity which represents the spectrum of the solution. When its value is stated, it must be stated for a particular wavelength (e.g. ϵ_{532}). Thus absorption spectroscopy can be used to quantify the amount of chemical present in an unknown solution.

Instrumentation:

Instruments for measuring the absorption of UV or visible radiation are made up of the following components;

1. Sources (UV and visible)
2. Wavelength selector (monochromator)
3. Sample containers
4. Detector
5. Signal processor and readout

Solutions:

Stock $\text{Cr}_2\text{O}_7^{2-}$ solution. Prepare 100 ml of 0.002 M of stock $\text{Cr}_2\text{O}_7^{2-}$ solution in 0.03 M H_2SO_4 .

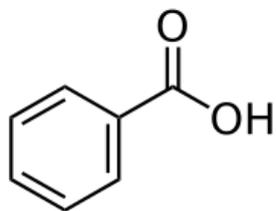
Stock MnO_4^- solution. Prepare 100 ml of 0.002 M of stock MnO_4^- solution in water.

Procedure

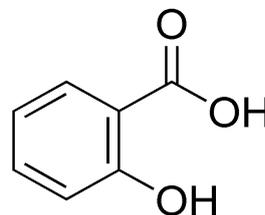
- 1) Prepare blank (0.03 M H₂SO₄), 1x10⁻⁴ M, 2x10⁻⁴ M, 3x10⁻⁴ M and 4x10⁻⁴ M of Cr₂O₇²⁻ and blank, 1x10⁻⁴ M, 2x10⁻⁴ M, 3x10⁻⁴ M and 4x10⁻⁴ M of MnO₄⁻ standard solutions in 50 ml volumetric flasks. In addition, prepare a mixture of Cr₂O₇²⁻ and MnO₄⁻ with the concentrations of 2 x10⁻⁴ M each.
- 2) Scan 4x10⁻⁴ M of Cr₂O₇²⁻, 4x10⁻⁴ M and MnO₄⁻ solutions and the prepared mixture from 250 nm to 750 nm and obtain three spectra.
- 3) Determine three working wavelengths;
 - a. A wavelength at which MnO₄⁻ shows maximum absorbance
 - b. A wavelength at which Cr₂O₇²⁻ shows maximum absorbance
 - c. An intermediate wavelength between two maximum absorbances from **a** and **b**.
- 4) Measure the absorbances of MnO₄⁻ and Cr₂O₇²⁻ standard solutions at three wavelengths and then three draw calibration plots for each wavelength.
- 5) Determine molar absorptivities of MnO₄⁻ and Cr₂O₇²⁻ at these wavelengths.
- 6) Obtain your unknown mixture from your TA.
- 7) Measure absorbances of the unknown mixture at the wavelengths determined in **3**.
- 8) Calculate concentrations of MnO₄⁻ and Cr₂O₇²⁻ in the unknown mixture.

Infrared Determination of Benzoic Acid and Salicylic Acid

Purpose: In this experiment, molecular structures of benzoic acid and salicylic acid will be analyzed qualitatively using an infrared spectrometer.



Benzoic acid



Salicylic acid

Introduction:

Infrared absorption is associated with vibrational motions of molecules. Infrared spectroscopy can be used for the determination of organic compounds. Generally infrared spectrum of an organic compound provides a unique fingerprint which is distinguished from all other organic compounds. In this experiment, benzoic acid and salicylic acid are identified.

Spectra of solids that are not soluble in an infrared transparent solvent are obtained by special techniques. Producing pellets by grinding the sample with potassium bromide (KBr) powder is very common to obtain absorbance spectra. As alternatives, diffuse reflectance (DR) and attenuated total reflectance (ATR) accessories are used to obtain spectra without initial sample preparation.

Instrumentation:

An IR instrument contains a source of infrared radiation, a sample container which should be infrared transparent, a wavelength selecting device, a detector and a signal processor, consecutively.

Commercially there are three types of instruments for infrared absorption measurements. These are dispersive instruments, multiplex instruments and non-dispersive instruments.

A dispersive instrument has a monochromator with a grating element to disperse the radiation coming from the source into its wavelengths and it is used as a wavelength selecting device. It is mostly designed double-beam, that is, incoming IR radiation is split into two beams in order to pass through the reference and sample materials.

The most commonly used type of multiplex instruments is Fourier transform (FT) instruments. FT instruments don't have grating element to disperse the light. FT instruments are generally based on the Michelson interferometer. In an FT instrument, IR radiation coming from the source firstly passes through the sample, and then passes through the interferometer.

Non-dispersive instruments are filter or non-dispersive photometers. They are designed for quantitative analysis. Generally they are not complex, easy to use.

Chemicals:

KBr powder

Benzoic acid

Salicylic acid

Procedure

- 1) Weigh about 100 mg KBr powder, grind in a mortar, make pellet and collect background spectrum.
- 2) Weigh about 20 mg benzoic acid, mix with about 100 mg KBr powder, grind the mixture in the mortar, make pellet of benzoic acid. Then collect IR spectrum of benzoic acid.
- 3) Repeat step 2 for salicylic acid.
- 4) Install DR accessory.
- 5) Collect background spectrum using mirror disk.
- 6) Place sufficient benzoic acid into the sample cup and collect IR spectrum of benzoic acid.
- 7) Repeat step 6 for salicylic acid.
- 8) Install ATR accessory.
- 9) Collect background spectrum without placing anything on the sample compartment.
- 10) Place sufficient benzoic acid on the sample compartment and collect IR spectrum of benzoic acid.
- 11) Repeat step 10 for salicylic acid.
- 12) Interpret and compare the spectra and show the functional groups.

Suggested Readings for Project 2

- **Skoog D.A.; Holler F.J.; Nieman T.A. *Principles of Instrumental Analysis - 5th Edn.*; Thomson Brooks/Cole: Belmont, CA, USA, 1998.**

Spectrofluorometric Determination of Aluminum

Topics to be covered (page: 355-379, ch. 15)

- Theory of fluorescence and phosphorescence
- Emission and excitation spectra
- Instrumentation
- Applications of Fluorimetry

Determination of Fluoride in Toothpaste using Ion Selective Electrode (ISE)

- Need to determine fluoride in toothpaste
- Reference electrode
- Indicator electrode
- Ion selective electrodes
 - Fluoride ion selective electrode
- Interferences encountered

(page numbers: 571-572, 591-594, 602-603)

If you are using an earlier or newer edition of the book, you must find the related subjects in your textbook.

You can also find information on above topics from:

- **Robinson J.W.; Skelly Frame E.M.; Frame II G.M. *Undergraduate Instrumental Analysis -6th Edn.*; Marcel Dekker, NY, USA, 2005.**
- **Internet sources**

Spectrofluorometric Determination of Aluminum

Purpose: In this experiment, concentration of aluminum in a solution will be determined by fluorescence spectrophotometer. To achieve this, aluminum-morin complex will be formed which has fluorescent property.

Introduction:

Molecular fluorescence spectroscopy is based on the emission of light by molecules, which are excited to emit their characteristic spectra by exposure to UV light of specific wavelengths. The wavelength at which a molecule may be excited to emit is referred to as the excitation wavelength, and the spectrum emitted by the sample the emission spectrum.

In molecular fluorescence spectroscopy, the emitted spectrum is monitored at right angles to the exciting radiation.

The intensity of emission at a specific emission wavelength is proportional to the concentration of the fluorescent molecule. This relationship forms the basis of quantitative analysis with this technique.

Instrumentation:

Instrumentation used to measure fluorescence involves

- an excitation light source
- a device to select the excitation wavelength
- a sample holder
- a device to select the fluorescence wavelength to be monitored
- a photon detector
- readout device

Chemicals and Preparations of Solutions:

- 1) Al^{3+} stock solution (1000 ppm): AlCl_3 is dissolved in water (use a 100 ml volumetric flask) and pH is adjusted to 2.0 with HCl before reaching 100 ml.
- 2) Al^{3+} standard solution (10 ppm): 1.0 ml of stock solution is diluted with water in a 100 ml volumetric flask.
- 3) Morin solution: Morin hydrate is dissolved in ethanol. After filtration, 0.5% (w/w) solution is prepared.
- 4) Ethanol
- 5) Acetic acid / Sodium acetate buffer (0.1 M): 0.6 ml of acetic acid and 1.36 g of sodium acetate is dissolved in water and diluted to 100 ml in a volumetric flask.

Procedure

- 1) Place 0.0, 1.3, 2.5, 3.7, and 5.0 ml of 10 ppm Al^{3+} standard solution into 25 ml volumetric flasks separately.
- 2) Add 0.5 ml of morin solution, 14 ml of ethanol and 5 ml of buffer solution into the volumetric flasks. Then dilute them to 25 ml with water.
- 3) Repeat step 2 for the unknown solution. Remember dilution factor.
- 4) After preparation of solutions, wait for 30 minutes.
- 5) Obtain emission spectra of the solutions in the wavelength range of 480-600 nm and in excitation wavelength 465 nm.
- 6) Using maximum absorbance values, plot calibration curve and determine aluminum concentration of the unknown sample.

Determination of Fluoride in Toothpaste using Ion Selective Electrode (ISE)

Purpose: In this experiment you will use an ion selective electrode to measure fluoride concentrations in toothpaste.

Introduction: The fluoride selective electrode produces a potential across a LaF_3 , solid ion exchange phase. LaF_3 exhibits affinity toward F^- . With the exception of OH^- , it does not interact with other substances.

The fluoride ion electrode contains an internal reference electrode, an internal fluoride standard, and the LaF_3 ion exchange crystal. An external reference electrode must be used to perform the measurement. Assuming that a saturated calomel electrode is used as the external reference, the potentiometric cell may be represented as



Sources of error include fluoride ion activity changes due to the ionic strength of solution, temperature, which can affect the measured potential through the Nernst equation which governs the electrode potential response and through the several equilibria that fluoride may have with species present in solution, and substances that complex fluoride in solution.

The most important errors are due to competing chemical equilibria. For the active LaF_3 part of the electrode, the most important interfering species is hydroxide, OH^- . The hydroxide ion complexes with LaF_3 crystal itself, in the same fashion as does for F^- . Subsequently, the potential across the crystal will be a function of $[\text{OH}^-]$ and will interfere with the fluoride determination. Another pH dependent effect is due to the basicity of F^- . HF is effectively a weak acid and at pH less than 5, acid-base equilibrium will affect the concentration of F^- in solution. In order to overcome these problems, solutions needs to be buffered to a pH between 5 and 9 to control hydronium and hydroxide concentrations.

In general, other fluoride chemical equilibria may be active in solution. Fluoride forms complexes with many polyvalent cation species found in natural and processed waters. Si^{+4} , Fe^{+3} , and Al^{+3} all form complexes with fluoride and thus interfere with measurements. The degree of interference depends on the concentration of the complexing ion and that of the fluoride. The addition of a pH 5 buffer containing a strong chelating agent eliminates the pH and polyvalent cation complexation interferences.

Apparatus and Reagents:

1. Corning model 450 digital pH/ion meter, combination pH electrode, fluoride selective electrode, and magnetic mixer with Teflon-coated stirring bar.
2. CDTA (1,2-cyclohexylene-dinitrilo-tetraacetic acid), sodium fluoride, sodium chloride, glacial acetic acid, and sodium hydroxide

Solutions:

1. *5 M Sodium hydroxide:* Dissolve 20 g NaOH in distilled water, cool, and dilute to 100 mL.
2. *100 ppm F stock solution:* Dissolve 0.1105 g NaF in distilled water and dilute to 500 mL in volumetric flask.
3. *10 ppm F standard solution:* Dilute 10 mL 100ppm stock F solution to 100 mL with distilled water.
4. *pH 5.0-5.5 buffer solution:* Add 28.5 mL glacial acetic acid, 29 g NaCl, and 2 g CDTA to approximately 250 mL distilled water in a 500 mL beaker. Stir to dissolve and cool to room temperature. Calibrate the pH meter/combination pH electrode using pH standards. Using the combination pH electrode, adjust the pH of the buffer solution to between 5.0 and 5.5 with 5 M NaOH (about 75 mL will be required). Transfer solution to a 500 mL volumetric flask and dilute to the mark with distilled water.

Calibration:

Calibrate the electrode as follows. Prepare a series of fluoride standards using the 10 ppm F standard solution in the range of 0 to 2.00 mg/L by diluting appropriate volumes to 50.0 mL using the series below:

Volume of standard solution (mL)	Standards (mg/L)
0.00	0.00
1.00	0.20
2.00	0.40
3.00	0.60
4.00	0.80
6.00	1.20
8.00	1.60
10.00	2.00

Procedures:

Sample preparation:

Weigh 200 mg of toothpaste in a small weighing boat and transfer quantitatively to a 250 mL beaker and add 25 ml water. Boil the mixture for 2 min., cool, and transfer quantitatively to a 250 ml volumetric flask. Add 1.4625g of solid NaCl and dilute to the mark.

Measurement procedure:

Set up the pF meter by connecting the fluoride selective electrode. Readings can be taken in potential (mVolts). To prepare for measurement, place 15.0 mL of sample or standard and 15.0 mL of buffer solution in a 100 mL beaker. Place beaker on magnetic stirrer and mix at medium speed. Immerse electrodes in the stirred solution. When taking measurements, electrodes must remain in solution for at least 3 minutes and until the potential readings have stopped drifting.

Data Sheet

Standards (mg/L)	Potential Readings (mV)
0.00	
0.20	
0.40	
0.60	
0.80	
1.20	
1.60	
2.00	
Sample	

Calculations:

1. Plot potential measurement of fluoride standards against concentration on two-cycle semi-logarithmic graphic paper. Plot milligrams F⁻ per liter on the logarithmic axis (ordinate), with the lowest concentration at the bottom of the graph. Plot millivolts on the abscissa.
2. Read the corresponding fluoride concentration from the standard curve, from the potential measurement for each sample.
3. Calculate % F in the original toothpaste sample.

Suggested Readings for Project 3

- Skoog D.A.; Holler F.J.; Nieman T.A. *Principles of Instrumental Analysis - 6th Edn.*; Thomson Brooks/Cole: Belmont, CA, USA, 1998.

Determination of Copper in Tap Water by Flame Atomic Absorption Spectrometry

Topics to be covered

- Basic theory of atomic absorption
- Sample atomization techniques
- Atomic absorption instrumentation
- Sources
 - Hollow cathode lamp
- Sensitivity and limit of detection
- Calibration methods
- Standard Addition Method

(page numbers: 11-18, 230-240)

The Determination of Sodium, Potassium, and Calcium in Drinking Waters by ICP-Atomic Emission Spectrometry

Topics to be Covered

- Basic theory of atomic emission.
- ICP instruments and their operation.
- Inductively Coupled Plasma Atomic Emission Spectroscopy
 - Plasma generation
 - Sample introduction
 - Axial and radial plasma
- Benefits of a plasma versus a flame source.
- How does an ICP work ?

(page numbers: 254-264)

If you are using an earlier or newer edition of the book, you must find the related subjects in your textbook.

You can also find information on above topics from:

- **Robinson J.W.; Skelly Frame E.M.; Frame II G.M. *Undergraduate Instrumental Analysis -6th Edn.*; Marcel Dekker, NY, USA, 2005.**
- **Internet sources**

Determination of Copper in Tap Water by Flame Atomic Absorption Spectrometry

Purpose: In this experiment the amount of copper in tap water will be determined by flame atomic absorption spectrometry.

Introduction:

Atomic absorption spectrometry involves the absorption of radiant energy by neutral atoms in the gaseous state. Elements in the samples are atomized in flame or graphite atomizer, and each element absorbs characteristic wavelength-light discharged from cathode lamp. By measuring the absorbance of the light, quantitative analyses are carried out. The element being determined must be reduced to the elemental state, vaporized, and imposed in the beam of radiation from the source. This process is most frequently accomplished by drawing a solution of the sample, as a fine mist, into a flame. The flame thus serves a function analogous to that of the cell and solution in conventional absorption spectroscopy. The light beam from the source is passed directly through the flame.

Quantitative analysis is easily performed by measuring the radiation absorbed by the sample in the flame. Typically a calibration curve is first prepared by measuring the absorbance of a series of solutions of known concentration. The sample(s) is then measured under same conditions. The instrument analyzes inorganic elements in the samples at the range of ppm (mg/L) and/or ppb ($\mu\text{g/L}$) levels. About 65 elements can be determined by atomic absorption with sensitivities down to 0.1 mg/L in some cases.

Instrument:

Thermo-Elemental Solaar M atomic absorption spectrometer

Starting the system:

After turning the instrument on, plug in the appropriate hollow cathode lamp. Once initialization is complete, create a new method, using the method wizard. Select “Cookbook” method and press the **Next** button to view each of the parameters. Record these parameters in your lab notebook.

Press **Lamp** button to turn the lamp on. The lamp should begin to glow.

Optimization:

Aligning the Lamps:

Once the method has been loaded, ensure the required lamp position is selected with lamp selection located in the carousel position. Allow the lamp time to warm up (about 15 minutes).

Ensure nothing is in the optical path, and then press the **Optical Set up** button. When the optical set up is finished, the system status will return to **ONLINE**. The lamp is now aligned and ready for use.

Aligning the Burner:

Use a piece of white to locate the light path. Do this by placing the paper level on the top of the burner inside the grid window.

Place the card halfway along the burner slot. Position the card with the vertical line perpendicular to the slot, and adjust the burner height until the light beam falls within the target area.

Lighting the Flame:

Open the acetylene cylinder. The pressure should already be set to ~ 9 psi. Open the air compressor. Press the 'Flame on' button and keep it pressed until the flame ignites. Watch this through the window. If the flame does not ignite or goes out, wait 5 seconds and then try again. If this fails, see your TA. Check the gas regulator settings and readjust if necessary.

When the flame has stabilized, adjust the flame conditions as described in the next section.

Optimizing the Flame Signal:

Aspirate the blank and click on the **Autozero** button to perform an Instrument Zero. Aspirate a standard solution that will give an absorbance of at least 0.2. Watch the signal bar and adjust the impact bead position by gradually turning the impact bead adjustment screw first clockwise, then counter-clockwise to find the maximum absorbance.

THE SYSTEM IS NOW OPTIMIZED.

Calibrating the Method:

Open the **Results** window so that you can see your results as they are measured and the calibration window to see the calibration plot as it is built up from the standard solutions.

Running the analysis:

Click on the **Analyse** button to start the analysis. As the analysis proceeds, you will be prompted to aspirate the solution required for each stage. Aspirate each solution as you are prompted and click on **OK**. The solution will be measured and the result will appear on the **Results** window.

Shutting Down the System:

Aspirate pure solvent for 10 minutes.

Shut off acetylene supply at the gas cylinder.

Shut off air tank.

Turn off the flame.

Wait until the hissing sound stops, then turn off the Instrument.

Reagents:

1. 250 ml 100 mg/L Stock Solution of Copper (made by diluting the appropriate weight of copper sulfate pentahydrate, $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$)
2. 65 % HNO_3 solution.

Procedure:

Sample and Standards:

1. To seven labeled 50 mL volumetric flasks, prepare the following solutions from 100 mg/L stock solution of copper:
0 (Blank), 0.5, 1.0, 2.0, 5.0, 10.0, and 20.0 mg/L Cu standards (before making dilution add 0.5 ml of 65 % HNO_3 solution to each of the standard.)
2. Obtain your tap water from your TA and prepare standards by standard addition method in 1.0% HNO_3 . (+0, +0.5, +1.0, +2.0, +5.0, +10.0 mg/L in 50 mL volumetric flask)

Data Sheet

A) AAS Operating Parameters

Operating conditions for Atomic Absorption Spectrometer	
Name of the instrument parts	
Light source	
Background correction	
Wavelength selector	
Detector type	
Spectral line used for Cu determination	

B) Absorbance Readings

Aqueous Standards	
Standards (mg/L)	Absorbance
0	
0.5	
1.0	
2.0	
5.0	
10.0	
20.0	

Standard Addition	
Standards (mg/L)	Absorbance
+0	
+0.5	
+1.0	
+2.0	
+5.0	
+10.0	

Calculations:

Plot the calibration curve of Abs vs. concentration for both standard addition and aqueous standards in the graph (using the least-squares method.).

Calculate the Cu concentration in tap water sample.

The Determination of Sodium, Potassium, and Calcium in Drinking Waters by ICP-Atomic Emission Spectrometry

Purpose: In this experiment, three elements commonly found in drinking water (sodium, potassium and calcium) will be analyzed and determined. The ICP will be operated in the sequential-multielement mode.

Introduction:

Inductively coupled plasma (ICP) emission spectrometry is a technique that is used primarily for the determination of trace concentrations of metals. The goal of ICP is to get elements to emit characteristic wavelength specific light which can then be measured. In ICP, a conducting gaseous mixture, plasma, containing a significant concentration of cations and electrons is formed. In the context of spectrometry, plasma can be defined as a partially ionized gas with sufficiently high temperature to atomize, ionize, and excite most elements. Plasma sources, in general, have several benefits over sources used in emission spectroscopy, with the main advantage of being highly energetic atomization sources. ICP is a type of flame that reaches temperatures of (6000-10000 K), much higher than those reached in ordinary combustion flames. Therefore, the atomization in ICP is more complete and the signal is correspondingly enhanced that enables much lower detection limits to be achieved.

Instrumentation:

An Inductively Coupled Plasma system typically includes the following components:

- Sample introduction system (nebulizer)
- ICP torch
- High frequency generator
- Transfer optics and spectrometer
- Computer interface

An ICP requires that the elements which are to be analyzed be in solution. The nebulizer transforms the aqueous solution into an aerosol.

The basic set up of an ICP consists of three concentric tubes, most often made of silica. These tubes are termed outer, intermediate and inner loops which collectively make up the torch of the ICP. The torch is situated within a water cooled coil of a radio frequency (RF) generator. As flowing gases are introduced into the torch, the RF field is activated and the gas in the coil region is made electrically conductive. The sequence of events forms the plasma.

In order to prevent possible short-circuiting as well as meltdown, the plasma must be insulated from the rest of the instrument. Insulation is achieved by the concurrent flow of gasses through

the system. Three gases flow through the system - the outer gas, intermediate gas, and inner or carrier gas. The outer gas is typically Argon or Nitrogen. The purpose of the carrier gas is to convey the sample to the plasma. Argon is commonly used for both the intermediate gas and inner or carrier gas.

The light emitted by the atoms of an element in the ICP must be converted to an electrical signal that can be measured quantitatively. This is accomplished by resolving the light into its component radiation (diffraction grating) and then measuring the light intensity with a photomultiplier tube at the specific wavelength for each element line.

Experimental:

Preparation of Standard Solutions:

Main Stock: 50 mL of 50.0 ppm Na, Ca, K standard solution. Take 2.5 mL of 1000 ppm stock solution of the multielement into a 50 mL volumetric flask and dilute to exactly 50 mL.

Before making dilutions add sufficient HNO₃ solution to the standards to get a final concentration of 1% (v/v) HNO₃.

Std.1 (50ml of 0.5 ppm std): Take 0.5 ml of stock solution into a 50 ml volumetric flask and dilute to exactly 50 ml.

Std.2 (50 ml of 1.0 ppm standard solution): Take 1.0 ml of main stock solution into a 50 ml volumetric flask and dilute to exactly 50 ml.

Std.3 (50 ml of 2.0 ppm standard solution.): Take 2.0 ml of main stock solution into a 50 ml volumetric flask and dilute to exactly 50 ml.

Std.4 (50ml of 5.0 ppm std): Take 5.0 ml of stock solution into a 50 ml volumetric flask and dilute to exactly 50 ml.

Std.5 (50 ml of 10.0 ppm standard solution): Take 10.0 ml of main stock solution into a 50 ml volumetric flask and dilute to exactly 50 ml.

Blank: Prepare a 1% (v/v) HNO₃ solution.

Preparation of Samples:

Sample 1: For the determination of Na⁺ only, dilute the sample 10 times

Sample 2: For the determination of Ca²⁺ and K⁺, use the sample undiluted

Note: Samples are also should be in 1 % (v/v) HNO₃.

Data Sheet

A) ICP-AES Operating Parameters under Optimized Conditions

Operating conditions for ICP-AES			
Name of the instrument parts		Detector voltage (V)	
Wavelength selector		Plasma gas flow rate (L/min)	
Detector type		Auxiliary gas flow rate (L/min)	
Spectral lines used for Na		Integration time (s)	
Spectral lines used for K		Pump flow rate (rpm)	
Spectral lines used for Ca		RF Power	

B) Intensity Readings of Na (I) 588.995 nm at different Instrumental conditions

i) Plasma Gas Flow rate and Pump flow rate is constant

Detector voltage, V	Intensity
1.	
2.	
3.	

ii) Detector Voltage and Pump flow rate is constant

Plasma gas flow rate, L/min	Intensity
1.	
2.	
3.	

iii) Detector Voltage and Plasma gas flow rate is constant

Pump flow rate , rpm	Intensity
1.	
2.	
3.	

C) Intensity Readings of standards and samples

Standards & Samples	Intensity Readings					
	Na (I)	Na (I)	Ca (I)	Ca (II)	K (I)	K (I)
Blank						
Std. 1						
Std. 2						
Std. 3						
Std. 4						
Std. 5						
Sample 1						
Sample 2						

Standards & Samples	Corrected Intensity Readings					
	Na (I)	Na(I)	Ca (I)	Ca(II)	K(I)	K(I)
Blank						
Std. 1						
Std. 2						
Std. 3						
Std. 4						
Std. 5						
Sample 1						
Sample 2						

Calculations & Discussions:

- Calculate the concentration of Na^+ , Ca^{2+} and K^+ in the water from some calibration graphs are obtained at the end of the experiment.
- Discuss the effect of detector voltage, pump flow rate and plasma gas flow rate on signal intensity
- Explain why the intensity of different emission wavelength for the same element is different from each other.

Suggested Readings for Project 4

- Skoog D.A.; Holler F.J.; Nieman T.A. *Principles of Instrumental Analysis - 5th Edn.*; Thomson Brooks/Cole: Belmont, CA, USA, 1998.

- Determination of o-Xylene, p-Xylene and Toluene by GC and GC-MS (page numbers: 701-716, 498-532).
- Determination of Caffeine Content in Normal and Diet Cola Samples by HPLC (page numbers: 725-760).

If you are using an earlier or newer edition of the book, you must find the related subjects in your textbook.

- Robinson J.W.; Skelly Frame E.M.; Frame II G.M. *Undergraduate Instrumental Analysis -6th Edn.*; Marcel Dekker, NY, USA, 2005.

Basics of Chromatography

- Know how a separation occurs in chromatography
- Be familiar with the terminology of chromatography. Some of the terms you should know are column, chromatogram, stationary phase, mobile phase, elution, eluent, eluate, solute, distribution constant, retention factor, separation factor, efficiency, band broadening, theoretical plate, and resolution
- Know all the different variables that affect the column efficiency
- Know the relationship between column efficiency and resolution
- Be familiar with the advantages and disadvantages of column chromatography
- Be able to explain how qualitative and quantitative information obtained from chromatography

Gas Chromatography

- Know the principle of separation in gas chromatography
- Be familiar with the basic instrumentation in gas chromatography and know each
- Know the advantages of capillary columns compared to packed columns
- Be able to select a specific detector and explain when to use that type of detector
- Be able to measure important quantities from a chromatogram (e.g. t_R , t_M , w , $w^{1/2}$)

Liquid Chromatography

- Be familiar with the different types of liquid chromatography
- Know what column band broadening is and how it can be minimized
- Understand the difference between normal phase and reverse phase partition chromatography
- Know typical mobile and stationary phases, and be able to predict elution order for a series of compounds
- Be able to explain how eluent strength affects the separation
- Be able to distinguish between gradient and isocratic elution
- Explain the differences between liquid and gas chromatography

Determination of Ortho-xylene, Para-xylene and toluene by GC and GC-MS

Introduction:

Toluene and the xylenes all belong to a group of organic compounds known as alkyl benzenes. The primary source of these alkyl benzenes in the environment is the petroleum industry and, to a lesser extent, the coke industry. Toluene and the xylenes are all used extensively as solvents and as raw materials in the synthesis of a variety of chemicals. All are used to some extent in the rubber and plastics industries, and all have been used as gasoline additives.

The alkyl benzenes are recognized primarily as atmospheric pollutants mainly because of their high volatility, but small amounts may enter aquatic and terrestrial systems (e.g., gasoline spills from leaking storage tanks).

In this experiment you will analyze an unknown sample containing o-xylene, p-xylene and toluene by gas chromatography (GC). It is one of the most popular chromatographic methods commonly used for the separation of non-polar compounds with high vapor pressure and a vaporization without decomposition.

Roughly, a gas chromatograph is composed from the carrier gas supply, the sample inlet, the column positioned in a column oven and the detector.

When a sample is injected into the column, the carrier gas transports the vaporized sample through the thermostated capillary column into the detector. As the sample mixture moves through the column, sample components that interact strongly with the stationary phase spend more time in the stationary phase vs. the moving gas phase and thus require more time to move through the column. When a substance leaves the column, it is sensed by a detector and the separated analytes are obtained as peaks in a chromatogram.

Reagents:

1. Xylene mixture (o-xylene + p-xylene)
2. Toluene
3. Dichloromethane
4. Syringe (10 μ L)

Solutions:

Preparation of o-xylene, p-xylene and toluene standards

Std 1 : 200 μ L **o-xylene** in 5 mL dichloromethane.

Std 2 : 200 μ L **p-xylene** in 5 mL dichloromethane

Std 3 : 200 μ L **toluene** in 5 mL dichloromethane

Procedure:

1. Set the instrument to optimum operating conditions for the determination of o-xylene, p-xylene and toluene mixture. Then, record all the information given on your data sheet.
2. Rinse the syringe with at least 3 volumes of your solution and dispense the rinses into the waste beaker.

As you take your sample press the needle through the septum of the injector.

Inject rapidly and wait for a few seconds again before withdrawing the needle.

Do not touch the injector head. It is HOT!

3. Inject 1.0 μL of o-xylene at 80 $^{\circ}\text{C}$ and 140 $^{\circ}\text{C}$ respectively and determine its retention time.
4. Inject 1.0 μL of p-xylene at 80 $^{\circ}\text{C}$ and 140 $^{\circ}\text{C}$ respectively and determine its retention time.
5. Inject 1.0 μL of toluene at 80 $^{\circ}\text{C}$ and 140 $^{\circ}\text{C}$ respectively and determine its retention time.
6. Inject 1.0 μL of o-xylene, p-xylene and toluene mixture into the gas chromatograph under the same conditions used for the standards.
7. Print the chromatogram of o-xylene, p-xylene and toluene standards.
8. Analyze the same mixture at 60 $^{\circ}\text{C}$ by GC-MS.

Data Sheet

A) GC operating parameters

GC Conditions for the determination of toluene, o-xylene and p-xylene	
Name of the instrument parts	
Carrier gas	
Injector temperature	
Column temperatures	
Detector temperature	
Column type	
Detector type	
Flow rate	
Sample size	

B) Properties of Compounds in Sample

Compound	Boiling Point	Molecular weight	Chemical Formula
Toluene			
o-xylene			
p-xylene			

C) Data for standards at T₁ (attach your chromatograms)

Compound	Peak #	Retention Time	Peak Resolution at °C	Toluene	o-xylene	p-xylene
Toluene				x		
o-xylene				x	x	
p-xylene				x	x	x

E) Data for standards at T₂ (attach your chromatograms)

Compound	Peak #	Retention Time	Peak Resolution at °C	Toluene	o-xylene	p-xylene
Toluene				x		
o-xylene				x	x	
p-xylene				x	x	x

Calculations:

Make your calculations on a separate sheet in your lab report based on the instructions given in the Analysis of Data part.

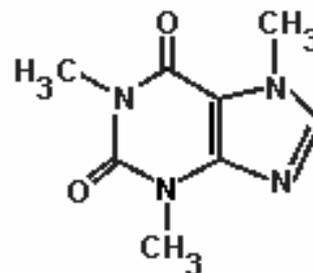
Analysis of Data:

1. Use the toluene, o-xylene and p-xylene standards to identify the each peak in your sample.
2. Record retention time of toluene, o-xylene and p-xylene standards on your data sheet.
3. Identify each peak in the chromatogram obtained.
4. Calculate the peak resolution of each standard relative to each other.
5. Comment on the results according to your data and calculations.
6. Do not forget to draw the block diagram of GC in your lab report and specify each component according to the instrument you have used.

Determination of Caffeine Content In Normal and Diet Cola Samples By HPLC

Introduction:

Caffeine is a common organic molecule found in many beverages such as coffee, tea, cola and some foods that we consume. Caffeine is a stimulant to the central nervous system. That is why many people drink coffee or soda to help them feel alert.



Caffeine has a mildly addictive effect on the body. When taken in small amounts caffeine is harmless, it is even considered beneficial; although, when taken excessively or in large doses for an extended period of time it has been proven to be detrimental. Therefore, it is interesting to know exactly how much caffeine is present in certain beverages.

This experiment provides an introduction to the application of High Performance Liquid Chromatography (HPLC) since it is the one way of determining the amount of caffeine present in these beverages.

HPLC utilizes a liquid mobile phase to separate the components of a mixture. These components (or analytes) are first dissolved in a solvent, and then forced to flow through a chromatographic column under a high pressure. In the column, the mixture is resolved into its components.

The main components of an HPLC system are a high-pressure pump, a column and an injector system as well as a detector. The system works as follows: eluent is filtered and pumped through a chromatographic column, the sample is loaded and injected onto the column and the effluent is monitored using a detector and recorded as peaks.

If a series of caffeine standards are analyzed, then the amount of caffeine in another substance can be determined. In this lab, you will prepare standard solutions of known caffeine concentration and find the caffeine content of some common cola samples.

Reagents:

1. Acetic acid (99.8%)
2. Sodium acetate
3. Acetonitrile
4. Caffeine

Solutions:

Preparation of mobile phase

1. 10% Acetonitrile: 90% Buffer solution ($\text{CH}_3\text{COOH}-\text{CH}_3\text{COONa}$).
2. Prepare a 0.02 M of 1L CH_3COONa solution.
3. Add CH_3COOH drop by drop until the pH value is 4.3, then mix 900 mL of buffer solution thus prepared with 100 mL acetonitrile.

Preparation of caffeine standards and samples

1. Prepare 100 ppm of 100 mL caffeine stock solution, then prepare 5 ppm- 10 ppm- 15 ppm- 20 ppm of standard caffeine solutions from 100 ppm stock.
2. Obtain a beverage sample.
3. For cola samples, degas the sample by placing it in an ultrasonic bath. Keep it until no more CO_2 bubbles appear in the sample.

Procedure:

1. Set the instrument to optimum operating conditions for the determination of caffeine. Then, record all the information given on your data sheet.
2. Rinse the syringe with at least 3 volumes of your solution and dispense the rinses into the waste beaker.
3. Inject about 20-25 μL of each standard caffeine solution or sample.
4. Switch the manual injector to the "load" position and push at least three 25- μL volumes of each sample through the sample loop, making certain each volume does not contain air bubbles.
5. With the syringe still in the injector, turn to the "inject" position, at which time the instrument will automatically start data acquisition. Leave the injector in the "inject" position, and remove the syringe.
6. Print the chromatogram of each standard caffeine solution.

Data Sheet

A) HPLC operating parameters

HPLC Conditions for the determination of caffeine in cola samples	
Name of the instrument parts	
Column type	
Detector type	
Mobile phase	
Flow rate	
Loop size	

B) Data for standards (attach your calibration curve)

Standards	Retention Time	Peak Area

C) Caffeine determination in cola samples

Beverage	Retention Time	Peak Area	Concentration	% Error

Calculations:

Make your calculations on a separate sheet in your lab report based on the instructions given in the Analysis of Data part and determine the caffeine content in your samples.

Analysis of Data:

7. Use the caffeine standards to identify the caffeine peak.
8. Record retention time and peak area for each of the standards on your data sheet.
9. Prepare a *calibration curve* by graphing peak area versus concentration of caffeine in ppm using the data obtained for the standard solutions.
10. Determine the amount of caffeine in cola samples. Report your results as instructed by your TA.
11. Use Table 1 to calculate the percent error between the results obtained in this experiment and the approximate amount of caffeine in your selected beverage.
12. Do not forget to draw the block diagram of HPLC in your lab report and specify each component according to the instrument you have used.

Table 1 – Approximate guide for caffeine amounts in common beverages.¹

Beverage	Caffeine (mg/mL)
Coca Cola	0.128
Diet Coke	0.128
Pepsi	0.105
Diet Pepsi	0.100
Tea, Iced	0.197
Tea, brewed, imported	0.290
Tea, brewed, domestic	0.193
Tea, instant	0.145
Drip Coffee	0.652
Espresso	1.690–2.25

¹National Soft Drink Association and Bunker and McWilliams, J. Am. Diet, 74:28-32, 1979.